Infiltration of tobacco leaf tissue

Introduction

- For transient expression in tobacco (*N. tobacum* and *N. benthamiana*) leaf lower epidermal cells
- Adapted from: Sparkes I, Runions J, Kearnes A and Hawes C., (2006), "Rapid transient expression of fluorescent fusion proteins in tobacco plants and generation of stably transformed plants", Nature Protocols, 1, (4), 2019-2025

Materials

- 4-6 week old *Nicotiana tobacum* or *Nicotiana benthamiana* plants
- Overnight culture (approx. 16h) of *Agrobacterium tumefaciens* (strains GV3101, EHA105 or ABI) containing binary plasmid

Method

- 1. make infiltration buffer (per sample approx. 3ml)
- 2. take 1ml of cells into 1.5ml Eppendorf tube and spin down in bench top centrifuge for 5 min at 5000rpm (or 3min at 8000rpm)
- 3. remove supernatant and add 1ml infiltration buffer, gently re-suspend pellet with pipette and repeat centrifugation to wash sample
- 4. remove supernatant and add 1ml of infiltration buffer
- 5. measure optical density of sample with spectrophotometer, take reading at 600nm absorption
- 6. calculate volume of sample to be used using formula Ci x Vi = Cf x Vf, where Ci is the optical density (OD), Cf is infiltration OD and Vf is $1000\mu l$; infiltration OD varies for each construct, usually between 0.01 and 0.1
- 7. pierce small hole into plant leaf and draw up resuspended cells into a 1ml syringe
- 8. place finger over hole, gently turn leaf to infiltrate the lower face of the leaf and gently push liquid sample into leaf
- 9. incubate for 2-3 days to allow proteins to be expressed before using infiltrated section for EM/confocal/protein extraction

Infiltration buffer

• stock buffer:

0.5M MES (store at 4°C)

0.02M Na₃PO₄.12H₂O (Sodium Perchlorate) (store at 4°C)

1M acetosyringone (in DMSO) (store at -20°C)

• 10ml infiltration buffer contain:

1ml 0.5M MES

1ml 0.02M Na₃PO₄.12H₂O

1µl 1M acetosyringone

0.05g glucose

Deionised water to make up total volume