

1 Integrating three comprehensive datasets shows that mitochondrial DNA variation is linked to  
2 species traits and palaeogeographic events in European butterflies.

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## 33    **Abstract**

34    Understanding the dynamics of biodiversity, including the spatial distribution of genetic diversity,  
35    is critical for predicting responses to environmental changes and for effective conservation  
36    measures. This task requires tracking changes in biodiversity at large spatial scales and correlating  
37    with species functional traits. We provide three comprehensive resources to understand the  
38    determinants for mitochondrial DNA differentiation represented by i) 15,609 COI sequences and ii)  
39    14 traits belonging to 307 butterfly species occurring in Western-Central Europe and iii) the first  
40    multi-locus phylogenetic tree of all European butterfly species. By applying phylogenetic  
41    regressions we show that mitochondrial DNA spatial differentiation (as measured with  $G_{st}$ ,  $G'_{st}$ ,  $D$   
42    and  $D_{st}$ ) is correlated with species traits determining dispersal capability and colonization ability.  
43    Due to the high spatial resolution of the COI data, we also provide the first zoogeographic  
44    regionalization maps based on intraspecific genetic variation. The overall pattern obtained by  
45    averaging the spatial differentiation of all Western-Central European butterflies shows that the  
46    paradigm of long-term glacial isolation followed by rapid pulses of post-glacial expansion has been  
47    a pervasive phenomenon in European butterflies. The results and the extensive datasets we provide  
48    here constitute the basis for genetically-informed conservation plans for a charismatic group in a  
49    continent where flying insects are under alarming decline.

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## 54    **Introduction**

55    Genetic diversity within populations and its spatial differentiation among populations are central  
56    concepts in biology. Within population diversity provides opportunities for populations to respond to  
57    shifting ecological pressures and inter-population differentiation triggers the processes of allopatric  
58    speciation (Coyne & Orr, 2004; Hughes, Inouye, Johnson, Underwood, & Vellend, 2008).  
59    Understanding the emergence and maintenance of genetic differentiation exposes fundamental  
60    evolutionary processes over a range of spatial and temporal scales. The resolution of such studies has  
61    advanced greatly since the onset of DNA sequencing (Allio, Donega, Galtier, & Nabholz, 2017;  
62    Bazin, Glémin, & Galtier, 2006; Lewontin, 1974; Nabholz, Mauffrey, Bazin, Galtier, & Glemin,  
63    2008).

64    A substantial effort has been devoted to verifying the prediction that neutral genetic diversity should  
65    equate to the product of mutation rate and effective population size. Despite this clear theoretical  
66    statement, DNA polymorphism appeared to be weakly correlated to population size and, when  
67    correlations have been found, the genetic diversity revealed is orders of magnitude smaller than  
68    expected based on differences in population size (Bazin et al., 2006; Leffler et al., 2012; Nabholz et  
69    al., 2008; Romiguier et al., 2014). Moreover, the results greatly varied among studies comparing  
70    genetic diversity for different taxa as well as when using different genetic markers (such as allozymes,  
71    nuclear or mitochondrial markers) (Allio et al., 2017; Bazin et al., 2006; Fujisawa, Vogler, &  
72    Barraclough, 2015; Leffler et al., 2012; Nabholz, Glémin, & Galtier, 2009; Nabholz et al., 2008;  
73    Romiguier et al., 2014).

74    As a major example, differentiation in nuclear (nDNA) and mitochondrial DNA (mtDNA) is expected  
75    to show different determinants even in the same model organisms. First of all, mtDNA has a faster  
76    mutation rate compared to nDNA and can show signatures of recent differentiation (e.g. intraspecific)  
77    as well as relatively old (Avise, 2009; Hebert, Cywinska, Ball, & deWaard, 2003). Secondly, because  
78    mtDNA is haploid, maternally inherited and recombination is limited to rare cases of heteroplasmy,

79 its effective population size is four times smaller and coalescence times shorter than in nuclear DNA  
80 (Allio et al., 2017; Nabholz et al., 2009). mtDNA is involved in respiration processes and has been  
81 found to be under strong selection (Galtier, Nabholz, GléMin, & Hurst, 2009; Nabholz et al., 2009;  
82 Pentinsaari, Salmela, Mutanen, & Roslin, 2016). Finally, mtDNA differentiation can be influenced  
83 by infections of microorganisms like *Wolbachia* (Galtier et al., 2009; Smith et al., 2012; Werren,  
84 Baldo, & Clark, 2008). Selection for variants determining different respiration performance and  
85 improved fitness in association with microorganisms, associated with high potential for genetic  
86 hitchhiking in the non-recombinant mitochondrial genome, make it difficult to disentangle neutral  
87 from adaptive mutations (Gillespie, 2000, 2001). Finally, population size may rapidly vary in  
88 geological time following environmental perturbations. It is thus expected that current effective  
89 population size and other species traits are poor predictors for the assumed consistent mutation rates  
90 and the resulting mtDNA polymorphism as expected by the neutral theory (Nabholz et al., 2009;  
91 Romiguier et al., 2014).

92 While several studies searching for fingerprints of effective population size and other species traits  
93 on DNA polymorphism have been carried out through inter-specific comparisons (Allio et al., 2017;  
94 Bazin et al., 2006; Fujisawa et al., 2015; Leffler et al., 2012; Nabholz et al., 2009; Romiguier et al.,  
95 2014), there are very few comparative phylogeographic studies which adopted a spatially explicit  
96 framework (Burney & Brumfield, 2009; Dapporto et al., 2017; Moritz et al., 2009). facilitating  
97 understanding inter-population patterns of genetic diversity and its determinants. The primary  
98 challenge for phylogeography is to adequately map current genetic diversity to allow the testing  
99 hypotheses that explain such variation. Usually patterns are explained by relatively recent events,  
100 such as Quaternary climatic oscillations (Avise, 2009; Hewitt, 2004). The increase in  
101 phylogeographic studies of multiple taxa opens the door to comparative work, which adds a layer of  
102 complexity in searching for shared sources of interspecific patterns, particularly in relating  
103 intraspecific genetic variation to environmental features and species traits (Bowen et al., 2016;  
104 Papadopoulou & Knowles, 2016). The final goal of comparative phylogeography is to disentangle

105 deterministic historical/contemporary and biotic/abiotic processes that determine the detected  
106 diversity (Dawson, 2014; Papadopoulou & Knowles, 2016).

107 Because of the relatively faster rates of divergence and coalescence compared to nDNA, mtDNA is  
108 a primary marker to study the distribution of diversity at the intraspecific level, with an almost  
109 ubiquitous use in phylogeography (Avise, 2009; Avise et al., 1987). This is particularly true for the  
110 cytochrome c oxidase subunit I (COI), a section of which has become the standard DNA barcode for  
111 animals (Hebert et al., 2003). Currently, public DNA barcode libraries contain millions of sequences  
112 (Kress, García-Robledo, Uriarte, & Erickson, 2015; Ratnasingham & Hebert, 2007) and now allow  
113 unknown samples to be identified, often to species level. The accumulation of DNA barcode data for  
114 an increasing number of groups, and in particular European butterflies, in public repositories  
115 (GenBank, BOLD) generated by wide scale research surveys (Dapporto et al., 2017; Dincă et al.,  
116 2015; Hausmann et al., 2011; Huemer, Mutanen, Sefc, & Hebert, 2014) is now extensive.

117 Here, we provide a novel assessment of which species traits correlate with different layers of intra-  
118 specific mtDNA differentiation, providing an overview of comparative phylogeography in western  
119 European butterflies. We provide the first zoogeographic regionalization map based on intra-specific  
120 genetic variation at the subcontinental scale of an entire superfamily (Papilionoidea). Our analyses  
121 are based on three novel resources now available for future studies: i) a DNA barcode dataset for the  
122 307 butterfly species occurring in western Europe (15,609 COI sequences of which 5,380 sequences  
123 were new for this study) (Fig.1); b) a database of 14 species features including feeding,  
124 morphological, natural history and ecophysiological traits for each of the 307 barcoded species and  
125 c) a phylogenetic tree for all 496 European butterflies based on the mitochondrial gene COI and 13  
126 nuclear markers. By integrating these datasets, we test three main predictions about mtDNA genetic  
127 diversification and its spatial structure.

128 First, since high selection, absence of recombination, erratic mutation rate and stochastic variation in  
129 population size make overall mtDNA divergence highly unpredictable (Allio et al., 2017; Galtier et

130 al., 2009; Nabholz et al., 2009; Romiguier et al., 2014), we expect to find no correlations between  
131 mtDNA diversity (haplotype diversity) and species traits related to population size, dispersal  
132 capability, number of generations and climatic tolerance (prediction 1).

133 Second, a plethora of studies demonstrated that mtDNA shows strong differentiation among  
134 populations particularly in poorly dispersive species. Consequently, we predict that when spatial  
135 information and genetic variation are assessed together, as typically done with widely used indices of  
136 population differentiation (Whitlock, 2011), a relationship with species traits should emerge (Burney  
137 & Brumfield, 2009; Dapporto et al., 2017) (prediction 2).

138 Finally, the Quaternary history of Europe has been dominated by climatic pulses which rendered most  
139 of Northern and Central Europe unsuitable for many ectothermic species, which became restricted to  
140 the southern peninsulas (Iberia, Italy, Balkans) and Mediterranean islands (Hewitt, 2004; Schmitt,  
141 2007) during cold periods. These refugia were separated from each other by conspicuous physical  
142 barriers such as sea channels and mountain chains (mostly represented by the Alps and Pyrenees)  
143 (Fig. 2) (Hewitt, 2004; Schmitt, 2007). We predict that a zoogeographic regionalization based on the  
144 intraspecific COI variation in our dataset will produce diversity patterns coherent with those expected  
145 on the basis of theoretical, geomorphological and palaeoclimatic expectations, as well as with those  
146 obtained by comparing communities based on faunistic data (prediction 3).

147

## 148 **Materials and Methods**

### 149 *Sampling and dataset*

150 We gathered 15,609 COI sequences belonging to 307 species occurring in Western Europe (Spain,  
151 Portugal, Andorra, France, United Kingdom, Belgium, Germany, Italy, Switzerland, Austria,  
152 Sweden, Norway, Denmark, Belgium, Netherlands). 5,380 COI sequences have been generated for  
153 this study by using standard procedures (see Supplementary Methods and Results), and the rest

154 have been obtained from BOLD (<http://www.boldsystems.org/>) and GenBank (10,229). Sequences  
155 have been screened to verify that i) they had a length of at least 500 bp, ii) they were georeferenced  
156 and iii) they were assigned to the correct species. The recent check list of European butterflies  
157 (Wiemers et al., 2018) has been used as a reference for taxonomy, but a series of species sharing  
158 DNA barcodes according to previous studies (Dincă et al., 2015; Dincă, Zakharov, Hebert, & Vila,  
159 2011) have been merged into a single entity because they share mitochondrial history  
160 (Supplementary Methods and Results). Sequences have been grouped into spatial units as follows:  
161 islands have been treated as individual units and sequences for the European mainland have been  
162 divided into areas of 2.5x2.5 degrees of latitude and longitude, resulting in 123 spatial units with at  
163 least one sequence (Fig. 1).

164

#### 165 *Indices of genetic differentiation*

166 For species occurring in at least 4 spatial units and with a minimum of 15 sequences in total we  
167 calculated three indices of genetic differentiation. The first, haplotype diversity (Hd), was  
168 calculated as the average of p-distance matrices among unique haplotypes using the “nuc.div”  
169 function of the “pegas” R package. This index is only dependent on the degree of differentiation  
170 among haplotypes regardless of their spatial distribution and frequency and it is typically used to  
171 measure mtDNA polymorphism (Nabholz et al., 2009).

172 The second is, the absolute differentiation among populations (Nei, 1987), is given by:

$$173 \text{ Dst} = \text{Ht} - \text{Hs}$$

174 where Ht represents the average p-distances for all specimens of a given species, and Hs is the  
175 average of the intra-unit p-distances. Thus, Dst represents the average genetic differentiation among  
176 areas in p-distance units. Species showing a higher differentiation among haplotypes (high Ht) and  
177 a spatial segregation (low Hs) have a maximum value for this index. Negative Dst values (intra-area

178 differentiation higher than inter-area differentiation) can have different subtle meanings, but are  
 179 most often generated as artefacts due to relatively small sample sizes; usually they are set to zero  
 180 (Meirmans & Hedrick, 2011) and we applied this solution.

181 The third measure was the widely used standardized index of population differentiation (Nei, 1987)  
 182 defined as:

183  $G_{st} = D_{st}/H_t$

184 which represents the fraction of the total genetic differentiation encompassed by the differentiation  
 185 among areas (Nei, 1987). This index ranges from negative values to 1 (complete differentiation) and  
 186 is independent of the number of changes exhibited by the different haplotypes of a given species.  
 187 Negative values have been set to zero (see above). The use of  $G_{st}$  has been debated as a measure of  
 188 population diversification for extremely variable markers (which is usually not the case for  
 189 mitochondrial markers) as it tends to underestimate differentiation among populations and to  
 190 strongly depend on intra-population variability (Jost, 2008; Whitlock, 2011). For this reason, we  
 191 also applied both  $D$  and  $G'_{st}$  indices, which are less affected by high values of  $H_s$  (see  
 192 Supplementary Methods and Results for their formulation).

193 *Species traits*

194 For the selected species we gathered a series of traits (Dapporto et al., 2017) representing four  
 195 (morphology, feeding, life history and physiology) of the five groups identified by Moretti et al.  
 196 (2017) to cover the primary functions of invertebrates: a) trophic generalism (feeding trait), was  
 197 identified as i) the number of host plant genera reported in two literature sources (Table S2); b)  
 198 mobility measured by the ii) wingspan proxy morphological trait as indicated by Sekar (2012) and  
 199 assessed as the average of minimum and maximum wingspan reported for each species in Higgins  
 200 & Riley (1970); c) phenology (life history trait) identified as iii) the number of months during  
 201 which adults occur in Europe, iv and v) the first and the last month when adults fly, and vi)



202 voltinism, i.e. the maximum number of generations per year recorded in Europe (Tolman &  
 203 Lewington, 2008). We also included a series of variables describing d) the climatic preference and  
 204 tolerance (physiological trait) according to Schweiger, Harpke, Wiemers, & Settele (2014).  
 205 Although these climatic niche indices cannot be considered as functional traits (Moretti et al.,  
 206 2017), they are widely used as proxies for the traits responsible for eco-physiological responses to  
 207 climate (Dapporto et al., 2017; Devictor et al., 2012). The variables we included are: vii) mean  
 208 annual temperature, viii) mean annual precipitation, ix) standard deviation of mean temperature, x)  
 209 standard deviation of mean precipitation, xi) upper 95% confidence limit of temperature mean, and  
 210 xii) lower 95% confidence limit of precipitation mean. Although direct information about effective  
 211 population size for all species over the entire continent is unavailable, their occurrence in Europe is  
 212 well assessed and range size, calculated as xiii) the number of 30x30km squares occupied  
 213 (Schweiger et al., 2014), is used here as a proxy for population size for at least two reasons: 1) the  
 214 species showing wider distributions can be expected to have a higher total of individuals across  
 215 their range, 2) butterfly species with larger ranges also tend to have more numerous local  
 216 populations (Brändle, Öhlschläger, & Brandl, 2002). Another distributional trait has been included  
 217 as xiv) the maximum altitude to which a species lives in Europe (Table S3).

218 Butterfly traits are usually highly inter-correlated and can be reduced to factors by using ordination  
 219 methods (Dapporto et al., 2017; Middleton-Welling, Wade, Dennis, Dapporto, & Shreeve, 2018).  
 220 Principal Component Analysis was applied to life history and physiology traits using the R function  
 221 “rda” and the components with eigenvalues higher than one have been retained as variables.

222 As a reference phylogeny, we constructed a phylogenetic tree for all 496 species of European  
 223 butterflies based on 14 genes (1 mitochondrial and 13 nuclear). The complete alignment was made  
 224 with ClustalW as implemented in BioEdit 7.2.5 (Hall, 1999) and consisted of 496 sequences (one  
 225 for each species) with a total length of 15,741 sites, 5,214 of them parsimony-informative,  
 226 containing the following genes: COI (covering 496 species with a total length of 1,532 sites and a

mean number of 1,087 nucleotides per species), wingless (283/467/386), EF1 $\alpha$  (282/1,725/1,022), rPS5 (143/760/594), GAPDH (137/714/651), CAD (103/2,928/946), MDH (67/750/590), IDH (65/710/681), H3 (57/329/328), RpS2 (42/862/454), DDC (27/2,012/689), HCL (21/633/623), Thiolase (21/1,020/1,020), and CAT (20/1,299/1,292). A maximum likelihood tree was estimated with IQ-TREE using the above alignment partitioned by genes and codon positions, with the substitution model option set to “Auto”, applying the FreeRate model with 4 rate categories, and default settings for branch support analysis and search parameters. The existence of a phylogenetic signal for species traits, environmental constraints and for Hd, Dst, D, G'st and Gst was tested with Pagel's lambda index by applying the “phylosig” R function of the "phytools" package.

## *Assessing predictions 1 and 2: Determinants for mtDNA differentiation*

The relationships between species traits and their Hd, Dst, D, Gst and G'st were assessed using phylogenetic regression. We used Pagel's lambda as a model for the phylogenetic covariance of residuals as implemented in the function “phylolm” of the R package “phylolm”. To avoid model overfitting and to provide a better parameterization of variables, we used the framework of multi-model inference of Generalized Linear Models through Information-Theoretic Approach (Burnham & Anderson, 2002) to select a set of “best models” by using the “MuMIn” R package. This approach allows selection of the best combination of predictors from the global model including all possible combinations. The model comparisons were performed adopting the corrected Akaike Information Criterion (AICc), and the model choice was done based on  $\Delta AICc$  (which represents the difference between each model and the most parsimonious model). We selected all models with  $\Delta AICc$  values  $< 4$ , considered to be equally parsimonious (Burnham & Anderson, 2002). According to this procedure only a small subset of predictors is selected as significantly affecting the response variable. The correlation coefficients of each predictor are averaged among the selected best-fitting models. The significance of the estimated coefficient is calculated with a z Wald test.

252

253 *Assessing prediction 3: Overall phylogeographic structure*

254 To provide a zoogeographic regionalization of South-Western Europe based on intraspecific  
255 diversification of COI sequences we applied the most recent procedures used in zoological  
256 regionalization based on a combination of hierarchical tree analysis to define clusters and  
257 unconstrained ordination to describe their relationships (Holt et al., 2013). At the basis of the  
258 procedure, a distance matrix among units was produced using pairwise  $G_{ST}$  among pairs of units for  
259 each species, using the following formula:

260  $G_{ST_{i,j}} = D_{ST_{i,j}} / H_t$

261 This represents the fraction of the overall genetic differentiation ( $H_t$ ) expressed by the population  
262 differentiation between a given pair of units (i and j).

263 Using the  $G_{ST}$  pairwise matrices for each species, we then calculated the mean of the available  
264 values of the corresponding cells of the matrix. We retained all the units that shared at least 10  
265 species to produce a final mean  $G_{ST}$  matrix, representing the degree of genetic differentiation among  
266 selected units based on all species. We then applied a Ward hierarchical clustering to this matrix.  
267 By using the “recluster” R package the tree was cut at different levels returning a series of  
268 clustering solutions. Then, a Principal Coordinates Analysis (PCoA) was applied to the dissimilarity  
269 matrix and we projected the configuration in the RGB space using the R package “recluster”  
270 (Dapporto et al., 2013). The colour resemblance of the resulting dots is proportional to the genetic  
271 similarity among the units. For each cut of the tree we attributed colours to the areas belonging to  
272 each cluster. These colours corresponded to the barycentre of area positioning in the RGB space.  
273 This “average colour” for each region has been used for mapping the zoogeographic regions as  
274 done by Holt et al. (2013).

275

## 276 **Results**

### 277 *Genetic dataset and species traits*

278 The 15,609 COI sequences belonging to 307 species have been grouped into 123 areas identified as  
279 islands or into areas of 2.5x2.5 degrees of latitude and longitude for the European mainland (Fig. 1).  
280 Among the 307 species for which at least one sequence was available, 224 fulfilled a minimum  
281 requirement set for the assessment of population diversification (at least 4 areas and 15 specimens).  
282 A full set of 14 traits describing feeding ecology, mobility, phenology, climatic tolerance and  
283 demography was available for 214 of these species. A PCA carried on four traits defining butterfly  
284 phenology identified only one component with an eigenvalue higher than 1 and it was mostly  
285 positively represented by voltinism and length of the flight period (Table S3, Fig. S1). For eco-  
286 physiological traits defining climatic tolerance, two components had eigenvalues >1 (Table S3, Fig.  
287 S2): the first component ordered species from those adapted to colder climates to those living in  
288 warmer areas; the second component ordered species mostly according to the precipitation they  
289 experienced (Table S3, Fig. S2). A phylogenetic tree was obtained by using 14 markers and all 496  
290 species of butterflies occurring in Europe (Wiemers et al., 2018) (Fig. 2) and showed an almost  
291 complete topological agreement with recent global phylogenies (Espeland et al., 2018). The tree  
292 allowed us to verify that the indices for population diversification (haplotype diversity, Dst, D, Gst,  
293 G'st) did not show any phylogenetic signal, while most functional traits did (Table 1).

294

### 295 *Prediction 1. mtDNA polymorphism and species traits*

296 As done in recent studies correlating genetic variability with species features (Fujisawa et al., 2015;  
297 Leffler et al., 2012; Romiguier et al., 2014), we performed phylogenetic regressions to model the  
298 three indices of genetic diversification against species trait controlling for phylogenetic signal.  
299 Phylogenetic regressions revealed that none of the selected traits significantly explained mtDNA

300 polymorphism measured as haplotype diversity (Table 2), although ecophysiology PC1, related to  
301 temperatures of the locations occupied was close to the significance threshold; species in current  
302 warmer areas tended to have higher mtDNA diversity than those of cold areas (Table 2).

303

#### 304 *Prediction 2. Population differentiation and species traits*

305 When the spatial information was added to the genetic differentiation among haplotypes in the  
306 assessment of Dst, phenology significantly explained the variation in overall population  
307 differentiation, with species characterized by longer flight periods and higher number of generations  
308 showing a lower level of differentiation (Fig. 3a, Table 3). An almost identical result was obtained  
309 by using D (Table S2) probably because for mtDNA the 1-Hs denominator tends to 1 due to the low  
310 intra-population differentiation in species showing spatial structure. Several species traits  
311 significantly correlated with Gst, only measuring the spatial segregation of haplotypes regardless of  
312 their degree of differentiation (Table 4). Species characterized by smaller wings and those  
313 exploiting a lower number of host plants (specialists), had higher population differentiation, with a  
314 similar, near significant, relationship for species with short flight periods. Widespread species also  
315 had high population differentiation (Table 4, Fig. 3 b-d). G'st showed very similar results to Gst  
316 (Table S1).

317

#### 318 *Prediction 3: Zoogeographic region with intra-specific differentiation*

319 Using the 224 species with sufficient data previously selected, we calculated a pairwise Gst matrix  
320 for each species among the areas where it has been found, and then an average Gst matrix has been  
321 calculated among areas. A Ward hierarchical clustering produced from the average pairwise Gst  
322 distance matrix was sliced at different levels as usually done for zoogeographic regionalization (Fig.  
323 4a). Due to higher species richness and a higher sampling effort in southern European regions, a  
324 subset of units having at least 10 shared species was concentrated in the Mediterranean area (Fig.

325 4b). The different hierarchical clustering solutions from  $K = 2$  to  $K = 6$  provided regionalization  
326 results that link nearby areas and that were highly coherent with existing geographic barriers (Fig.  
327 4b-f). The first node separated Iberia (except for Catalonia), the Balearics and Sardinia from the  
328 other areas (Fig. 3b). A solution with three clusters divided Sicily from the Pyrenees, Alps and  
329 Italian peninsula, according to a well-known efficient barrier to dispersal represented by the narrow  
330 Messina strait (Dapporto, Bruschini, Dincă, Vila, & Dennis, 2012; Vodă, Dapporto, Dincă, & Vila,  
331 2015) (Fig. 4c). The fourth cluster recognised Sardinia and the Balearics as a unit, according to a  
332 well-known and still largely unexplained similarity between these areas that may be explained by a  
333 refugium hypothesis (Dincă et al., 2011; Vodă et al., 2015) (Fig. 4d). The fifth cluster separated the  
334 Alps and Pyrenees from the Italian peninsula, with Corsica and circum-Italian islands resembling  
335 more the Pyrenees-Alps, reflecting a recurrent phylogeographic pattern found in several butterfly  
336 species (Dapporto et al., 2012) (Fig. 4e). The sixth cluster produced the expected division between  
337 the Alps and Pyrenees with Corsica, Elba and Giglio resembling more the Pyrenees than the  
338 spatially closer Alps (Dapporto et al., 2017) (Fig 4f).

339

## 340 **Discussion**

341 The three comprehensive resources (COI sequence dataset, species traits and phylogenetic tree)  
342 here presented for butterflies of Western Europe allowed us to test three specific predictions  
343 regarding mtDNA intraspecific genetic differentiation. First, we confirmed that overall intraspecific  
344 genetic variation in mtDNA (i.e. COI polymorphism) cannot be explained by any of the selected  
345 species traits, which were chosen to cover most of the main functions of invertebrates; dispersal,  
346 feeding, natural history, ecophysiology and distribution, the last being a proxy for population size  
347 (prediction 1). Second, when a spatially-explicit framework was applied, genetic differentiation  
348 among populations showed an effect for species traits, mostly when the influence of absolute  
349 genetic divergence is removed, as done by using  $G_{st}$  (and  $G'_{st}$ ). This indicates that the emergence

350 and maintenance of mtDNA differentiation is, at least in part, deterministically shaped by species  
351 ecology and by historical factors. Our result also supports the use of COI as a marker to understand  
352 ecological fingerprints in mtDNA spatial differentiation (prediction 2). Finally, the zoogeographic  
353 regionalization based on average COI population differentiation agreed with the main European  
354 biogeographical paradigm, which suggests that most butterfly species were isolated in restricted  
355 southern European areas during glacial periods, where they differentiated. From these refugia, the  
356 different lineages of most species experienced pulses of poleward expansion during the warmer  
357 interglacial periods, producing the observed recurrent suture zones along physical barriers (Alps,  
358 Pyrenees).

359

#### 360 *Prediction 1. mtDNA polymorphism and species traits*

361 Substitution rate and degree of polymorphism of mitochondrial and nuclear DNA are known to vary  
362 largely among taxa (Allio et al., 2017; Bazin et al., 2006; Fujisawa et al., 2015; Leffler et al., 2012;  
363 Nabholz et al., 2009; Romiguier et al., 2014; Welch, Bininda-Emonds, & Bromham, 2008). In  
364 general, animal groups show differences in DNA substitution rates that are linked to their population  
365 size or related traits (species range or body mass) or to other pressures acting on mtDNA (e.g.  
366 generation time, fecundity, homeo-heterothermic physiology) (Allio et al., 2017; Nabholz et al., 2009;  
367 Pentinsaari et al., 2016). On the other hand, there is mixed evidence for the expected relationship  
368 between intraspecific mtDNA polymorphism and species traits (Bazin et al., 2006; Fujisawa et al.,  
369 2015; Nabholz et al., 2009; Romiguier et al., 2014).

370 Despite the theoretical expectation that levels of neutral genetic variation should increase with  
371 effective population size, no relationships between range size (a likely good proxy for population  
372 size) and mitochondrial DNA diversity has emerged in our extensive dataset. Similarly, other  
373 species traits determining butterfly climatic and feeding generalism (ecophysiological traits and  
374 number of host plants), dispersal capability (length of flight period and wingspan) as well as the

375 number of generations are expected to influence the degree of gene flow and thus the level of  
376 genetic differentiation. Nevertheless, no significant relationships emerged between these traits and  
377 mitochondrial diversity. As previously suggested, the causes for the absence of these correlations  
378 likely reside in the non-neutral nature of mtDNA variation, in its apparently erratic mutation rate  
379 and in the strong fluctuations affecting both mtDNA polymorphism and population sizes in  
380 historical times (Bazin et al., 2006; Nabholz et al., 2009, 2008; Romiguier et al., 2014). The absence  
381 of any phylogenetic signal in mtDNA overall intraspecific genetic diversity, as well as in the indices  
382 of spatial differentiation, combined with most species traits being similar among closely related  
383 species supports the erratic behaviour of mtDNA polymorphism and its highly stochastic  
384 determinants.

385 mtDNA variants can determine different respiration performances (Toews, Mandic, Richards, &  
386 Irwin, 2014) and adaptive mutations are expected to rapidly spread in a population and even across  
387 populations, producing selective sweeps (Bazin et al., 2006; Galtier et al., 2009). Similarly, the  
388 maternally-inherited symbiotic bacterium *Wolbachia* can induce reproductive alterations (e.g.  
389 feminization, male killing, cytoplasmatic incompatibility) which are adaptive for the bacterium by  
390 enhancing the production of infected females (Werren et al., 2008). *Wolbachia* infection can result  
391 in a single haplotype (or haplogroup) dominating an entire population and there is growing evidence  
392 in European butterflies that different mtDNA lineages are associated with different infection status  
393 or strains of this bacterium (Dincă et al., 2018; Hernández-Roldán et al., 2016; Ritter et al., 2013).

394 Due to the non-recombinant transmission of mtDNA, genetic sweeps strongly affect the entire  
395 mtDNA genome, thus drastically lowering or eliminating former genetic diversity. This  
396 phenomenon occurs more frequently in species with a large effective population size, thus  
397 counterbalancing the emergence of a larger number of genetic variants (genetic draft) (Gillespie,  
398 2000, 2001). In-depth studies of mtDNA polymorphism in vertebrates have provided evidence that  
399 selective sweeps and genetic drift are less frequent than previously hypothesized (Allio et al., 2017;



400 Karl, Toonen, Grant, & Bowen, 2012), but this may not be the case in insects, where effective  
401 population sizes are orders of magnitude higher.

402 In addition, demographic stochasticity and historical changes in population size may be  
403 fundamental factors explaining the absence of a relationship between range size and mtDNA  
404 polymorphism in European butterflies. Nevertheless, due to the rarity of fossil data for butterflies,  
405 ranges and population sizes can only be calculated in the contemporary climatic conditions, which  
406 represent a warm interglacial period after a series of longer cold periods (Augustin et al., 2004).

407 Given the rapidity with which butterflies can shift their distribution tracking suitable climatic  
408 conditions with recent climate change (Devictor et al., 2012), it can be expected that most species  
409 now having restricted distribution on mountains were more widely distributed during cold periods  
410 of the Pleistocene, while many warm loving species were much more restricted to southern refugia,  
411 thus making contemporary population size uncorrelated with current mtDNA polymorphism  
412 (Nabholz et al., 2009). The fact that species now occurring in Mediterranean areas, which had likely  
413 been restricted to isolated southern refugia during the longer cold periods, showed a trend for higher  
414 mtDNA polymorphism could support this hypothesis. Finally, traits may not be fixed in time, but  
415 only their current states are known, and analyses using current traits may thus not fully reveal the  
416 historical relationships between mtDNA diversification and traits.

417

#### 418 *Prediction 2. Population differentiation and species traits*

419 Although the genetic variation of mtDNA can be wiped out by selective sweeps or mixed among  
420 populations after dispersal events in highly mobile taxa, most species show strong genetic structuring  
421 among more or less isolated populations, strongly supporting the use of mtDNA as a marker in  
422 phylogeography (Avice, 2000, 2009). Indeed, in case such events lead to the elimination of genetic  
423 differentiation, a spatial structure can be re-established in a relatively short time by the emergence of  
424 new haplotypes and lineages or by population dynamics such as gene surfing (Waters, 2011). For

example, a number of butterfly populations inhabiting Mediterranean islands that were connected to the mainland until the end of the last glacial period (about 15ka) show highly diverged mtDNA compared to populations inhabiting the mainland (Dapporto et al., 2017). This might be due to the recent occurrence of selective sweeps or shifts of lineages along the mainland (Mallet, 2010; Moritz et al., 2009), which have not yet reached insular populations due to sea barriers (Dapporto et al., 2017; Livraghi et al., 2018). These mechanisms could maintain spatially structured populations even if their overall degree of divergence strongly changes in time. In this case, we can expect that populations separated by the same semi-permeable barriers (mountain chains and relatively narrow sea channels) do not show homogeneous divergence times. This is a recurrent finding in comparative phylogeography as found e.g. in butterflies of the Tuscan Archipelago, Sardinia and Corsica (Dapporto et al., 2017), in Neotropical birds (Burney & Brumfield, 2009) and in many Australian taxa (Moritz et al., 2009).

When the spatial information is added to genetic differentiation, correlations between mtDNA differentiation and species traits emerged.  $D_{st}$  and  $D$  are still largely dependent on overall diversification being measured in terms of percentage divergence (Nei, 1987; Whitlock, 2011). The variation in these indices was significantly explained by phenology traits, with species characterized by a longer flight period and a higher number of generations showing a lower population differentiation. This is a highly expected result for butterflies, since the winged adults are the dispersive stage. Accordingly, species showing a shorter flight period have lower possibilities to cross physical barriers (Dapporto & Dennis, 2009), resulting in higher probabilities to diverge in allopatry, with longer times required to attain secondary sympatry among lineages and in a slower propagation of genetic sweeps.

Contrary to  $D_{st}$  and  $D$ ,  $G_{st}$  and  $G'_{st}$  are pure numbers that can reach the maximum value of one even if a single mtDNA substitution is completely segregated among populations. Compared to  $D$  and  $D_{st}$ ,  $G_{st}$  and  $G'_{st}$  showed a significant correlation with a higher number of species traits. Smaller species,

species exploiting less genera of food plants, and species with a larger range size showed a significantly higher COI genetic spatial structure – and species with a shorter flight period also showed a strong trend in the same direction. Range size is expected to correlate with genetic spatial structure since species with a larger range are expected to have more possibilities for divergence as barriers to gene flow can occur within their distribution and to thus comprise different lineages. In turn, wingspan is a well-known correlate of dispersal capability in butterflies (Dennis, Hardy, & Dapporto, 2012; Sekar, 2012). It has been found to correlate with genetic divergence and occupancy in different insular populations of European butterflies (Dennis et al., 2012). Similarly, exploiting a limited range of hostplants (specialism) may mean that resources are not generally widespread and thus dispersal is limited, promoting population segregation (Dennis et al., 2012).

This is in line with results obtained in comparative studies on birds, where tendency to secondary sympatry was positively correlated with a characteristic of wing morphology determining dispersal capabilities (Pigot & Tobias, 2014) and genetic differentiations across south American barriers correlated with life history traits (Burney & Brumfield, 2009). It must be noted that although traits had a significant effect on Dst and Gst and most traits show a clear phylogenetic association, there is no phylogenetic signal in any index we measured, as was found in birds (Burney & Brumfield, 2009). This could be because erratic mutation rate and genetic sweeps can produce strong contrasts between closely related taxa even when the processes generating and maintaining genetic structure are facilitated by species traits (Allio et al., 2017; Nabholz et al., 2009).

469

### 470 *Prediction 3. Zoogeographic region with intraspecific differentiation*

To the best of our knowledge, we here present the first zoogeographic regionalization at the sub-continental level based on intraspecific genetic diversification. Zoogeographic regions have been assessed so far by comparing faunistic communities, i.e. they were based on species distribution/occurrence differences (Holt et al., 2013). However, it is expected that a zoogeographic

475 assessment based on averaging intraspecific population differentiation among hundreds of species  
476 should i) reflect the main physical barriers and palaeogeographic history of the study area; and ii)  
477 correlate with a zoogeographic regionalization based on species composition, because the barriers  
478 separating species distributions are also expected to limit gene flow.

479 During the long Quaternary cold periods, most of the current European species were likely limited to  
480 the southern regions of the continent, represented by three peninsulas (Iberia, Italy, Balkans) and by  
481 several Mediterranean islands (Hewitt, 2000). Accordingly, the existence of different lineages of  
482 butterflies in these areas with narrow suture zones at the physical barriers among them (Alps,  
483 Pyrenees and sea channels), represents a pervasive pattern in European phylogeography (Bowen et  
484 al., 2016; Hewitt, 2000; Schmitt, 2007). According to this paradigm, we found evidence for six  
485 different zoogeographic regions that largely agree with the hypothesis of divergence in different  
486 Pleistocene refugia.

487 A similar regionalization was obtained by comparing butterfly communities in the same area  
488 (Dapporto, Fattorini, Vodă, Dincă, & Vila, 2014). The main difference between the assessment at  
489 community and intraspecific differentiation level refers to Sardinia, Corsica and the Balearic islands.  
490 These differences are rooted in the fundamental differences of the two assessments. In the analysis at  
491 the community level, the considerable number of endemic species from Sardinia and Corsica  
492 determined a high contrast and resulted in a highly distinct group; the Balearics without any endemic  
493 butterfly species, appeared very similar to Iberia (Dapporto et al., 2014). In the assessment of genetic  
494 diversity, the Sardo-Corsican endemics can only generate contrasts between Sardinia and Corsica  
495 (and Tuscan Islands) when they show distinct populations between these areas, as is often the case  
496 (Dapporto et al., 2014). Most of the pattern obtained with intraspecific genetic variation is instead  
497 encompassed by widespread species responsible for determining similarity/dissimilarity patterns  
498 among islands and mainland. Previous studies on butterflies showed that in many cases Sardinia and  
499 Corsica differ in their populations: those populations from Corsica and Tuscan Islands mostly

500 resemble those occurring in the Alps and the Pyrenees, whilst populations from Sardinia are often  
501 similar to those occurring in Iberia and the Balearics (e.g. *Callophrys rubi*, *Maniola jurtina*, *Pararge*  
502 *aegeria*, *Coenonympha pamphilus*) (Dapporto et al., 2017; Dincă et al., 2015; Livraghi et al., 2018).  
503 The analysis of intraspecific genetic divergence captured this main pattern, the determinants of which  
504 are still largely unexplained. The existence of distinct lineages from Iberia, Sicily and the Italian  
505 Peninsula is a very common pattern in butterfly phylogeography, while the diversification in the  
506 regions of the Alps and the Pyrenees is mainly determined by a different admixture of lineages (Dincă  
507 et al., 2018, 2015; Hernández-Roldán et al., 2016; Schmitt, 2007).

508

## 509 **Conclusion**

510 Among the challenges imposed by the current and accelerating biodiversity loss (Dirzo et al., 2014),  
511 understanding the dynamics of biodiversity is critical for predicting future scenarios and undertaking  
512 effective conservation measures (Hoffmann et al., 2015; Joly et al., 2014; Pacifici et al., 2015). This  
513 endeavour requires cheap, fast and reliable approaches to map and track changes of biological  
514 diversity over spatial and temporal scales, as well as linking them with species functional traits (Joly  
515 et al., 2014; Kress et al., 2015; Pacifici et al., 2015; Stein, Martinez, Stiles, Miller, & Zakharov, 2014).  
516 In the last decades, mitochondrial DNA has become increasingly prominent in biodiversity research,  
517 notably for phylogenetics, phylogeography and in the study of divergence processes (Avise, 2009;  
518 Burney & Brumfield, 2009; Cameron, 2014; Dincă et al., 2015; Galtier et al., 2009; Hernández-  
519 Roldán et al., 2016; Joly et al., 2014; Kress et al., 2015).

520 Nevertheless, the validity of many of the claimed advantages of using mtDNA as a marker in  
521 molecular ecology has been questioned in the last decade (Galtier et al., 2009; Stein et al., 2014). In  
522 fact, the assumption of neutrality of mtDNA has been weak and, in addition, mtDNA variation is not  
523 necessarily representative of genomic variation, as it is subjected to different determinants and  
524 inheritance mechanisms. We provide evidence that mtDNA spatial differentiation has a deterministic

525 fingerprint, being correlated with species traits known to determine the dispersal capability and  
526 colonization ability of butterflies (Dennis et al., 2012). Thus, we argue that mtDNA should be still  
527 considered as a fundamental marker for the understanding of ecological and evolutionary processes  
528 (such as demographic changes and dispersal patterns) that affect the historical and contemporary  
529 spatial ecology of species (or at least of their female populations). Moreover, the fall of the neutrality  
530 assumption for mtDNA implies the importance of mtDNA variation in influencing functional traits.  
531 Recent evidence proved that these mitochondrial-derived traits are involved, among others, in local  
532 adaptation to climatic conditions (Toews et al., 2014). Under this perspective, the strong variation in  
533 mtDNA sequences among populations and the relatively fast shifts in their distributions demonstrated  
534 by direct and indirect evidence, indicate that spatial mtDNA variation represents a source of  
535 differential adaptive optima, which can sweep across populations and preserve species in a rapidly  
536 changing environment (de Lafontaine, Napier, Petit, & Hu, 2018). The macroscopic consequence is  
537 the preservation of species diversity and of the related ecosystem functioning. In this vein,  
538 information about mtDNA variation has been recommended to be taken into consideration in  
539 conservation plans and reintroductions of butterflies (Dincă et al., 2018). The results and the extensive  
540 dataset provided here can constitute a basis to produce genetically informed conservation plans for a  
541 highly charismatic group in a continent where flying insects have been proven to be under incessant  
542 decline (Hallmann et al., 2017).

543

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753 **Author contributions**

754 LD designed the paper framework; LD, AC, RVo, VD, MM, LPC, SS, TS, RVi, collected the  
755 specimens, identified them at species level and carried out DNA sequencing and mined data from  
756 DNA repositories (BOLD and GenBank); LD, AC, GM, MM, EB, SB, LPC, gathered the trait dataset;  
757 MW gathered nuclear and mitochondrial markers and built the phylogenetic tree; LD carried out  
758 comparative phylogeography analyses; all authors participated in interpreting the results, in writing  
759 and editing the manuscript.

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761 **Data Accessibility**

762 - DNA sequences and specimen information: The DNA sequences, the information about collection  
763 data, taxonomy and GenBank and BOLD accession codes are available in the BOLD project DS-  
764 WEUP and from Dryad: doi: 10.5061/dryad.2q76p8f.

765 Butterfly traits are on Dryad: doi: 10.5061/dryad.2q76p8f

766 Phylogenetic tree: available on Dryad doi: 10.5061/dryad.2q76p8f

767 R scripts and file used to carry out the analyses available on Dryad: doi: 10.5061/dryad.2q76p8f



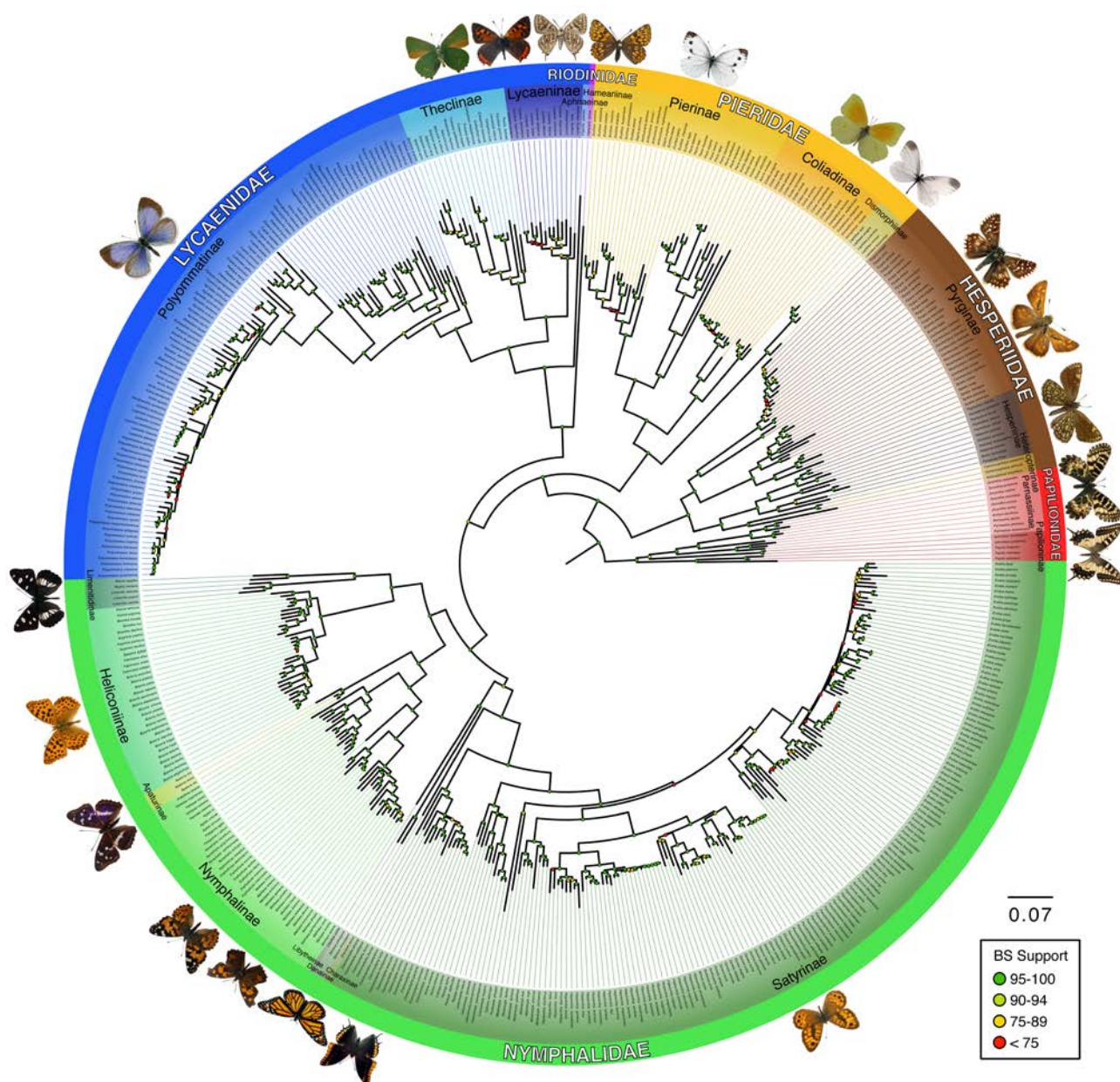


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770 Figure 1. Map of the study area and areas used for the assessment of spatial genetic variation. Dots  
 771 represent barycenters of sequences collapsed to squares of 2.5 degrees of latitude and longitude, and  
 772 to small islands, which are treated independently. The colour of the dots is proportional to the log of  
 773 the number of species analysed in each area (see legend). The mountain chains (Alps and Pyrenees)  
 774 separating the two main southern peninsulas (Iberian and Italian Peninsulas) are highlighted.

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778 Figure 2. The multi-locus phylogenetic tree for all European butterfly species. Families and  
 779 subfamilies are indicated as well as supports for nodes (BS support, in legend).

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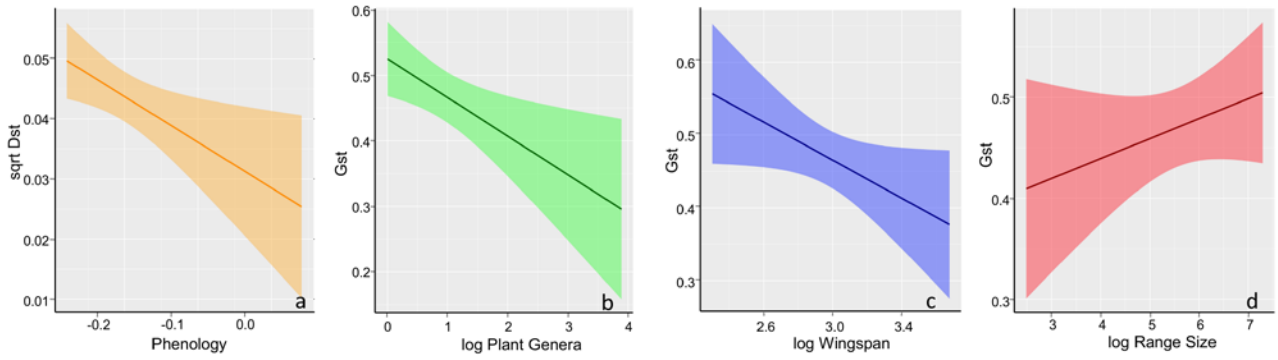


Figure 3. The significant univariate linear relationships between (a) square root-transformed Dst and phenology, (b) Gst and log-transformed number of plant genera used by species, (c) Gst and wingspan, and (d) Gst and range size. Shaded areas represent 95% confidence regions.



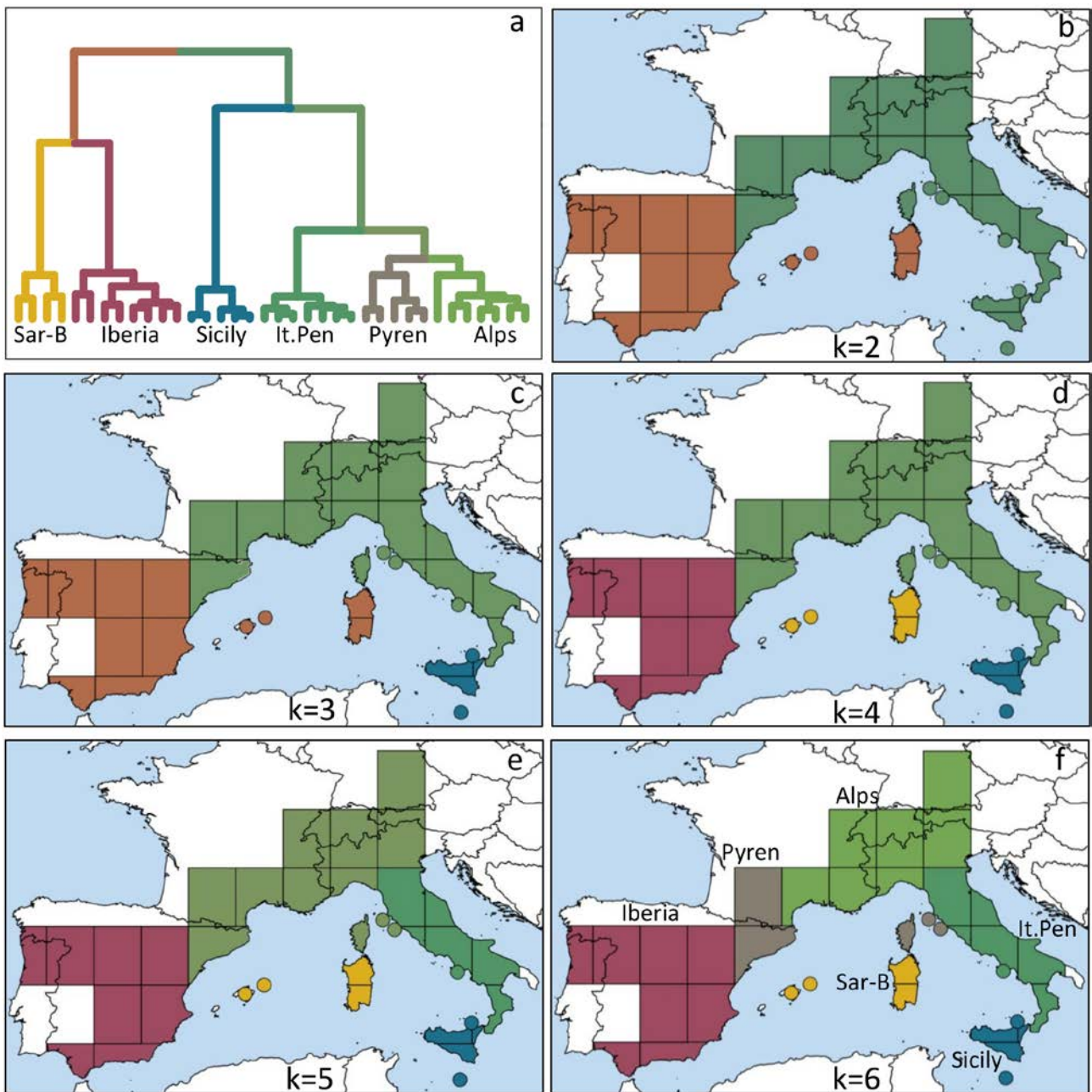


Figure 4. The tree obtained by applying the Ward algorithm to the average  $G_{ST}$  distances based on 226 species in the North-Western Mediterranean among a series of 36 areas (a). The tree is cut at different nodes to obtain five solutions from 2 to 6 clusters (b-f). For each solution, the tree branches are represented by using the colours obtained by projecting the bidimensional representation of the original dissimilarity matrix in RGB space and then by calculating the barycentres of the dots belonging to each subtree. The same colours are used in the maps to visualize the different zoogeographic regions. The regions identified by the solution for  $k=6$  are reported in the tree and in figure 3f; Iberia, Iberian Peninsula; Sar-B, Sardinia and Balearics; Sicily, Sicily and circum-Sicilian islands (Malta and Vulcano); It.Pen, Italian Peninsula and Capri; Pyren; Pyrenees, Corsica and surrounding islands (Elba and Giglio); Alps, Alps.

812 Table 1. Phylogenetic signal for the three indexes of COI genetic diversification and for the  
813 examined species traits. Variables highlighted in bold showed a significant effect.

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|                          | Pagel's<br>lambda | P                |
|--------------------------|-------------------|------------------|
| Gst                      | <0.0001           | 1.000            |
| G'st                     | <0.0001           | 1.000            |
| Dst                      | <0.0001           | 1.000            |
| D                        | <0.0001           | 1.000            |
| Nucleotide Diversity     | 0.021             | 0.550            |
| <b>PC1 phenology</b>     | <b>0.589</b>      | <b>&lt;0.001</b> |
| <b>Range size</b>        | <b>0.336</b>      | <b>&lt;0.001</b> |
| <b>PC2 ecophysiology</b> | <b>0.183</b>      | <b>&lt;0.001</b> |
| Max Altitude             | <0.0001           | 1.000            |
| <b>Host plants</b>       | <b>0.362</b>      | <b>&lt;0.001</b> |
| PC1 ecophysiology        | 0.011             | 0.710            |
| <b>Wing size</b>         | <b>0.988</b>      | <b>&lt;0.001</b> |

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818 Table 2. Conditional average results among the selected models in a phylogenetic regression  
819 comparing haplotype diversity with butterfly traits. Prediction 1. Variables highlighted in bold  
820 showed a significant effect.

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| HD                | Estimate | Std.Error | Z     | P     | models(29) |
|-------------------|----------|-----------|-------|-------|------------|
| PC1 ecophysiology | 0.175    | 0.092     | 1.894 | 0.058 | 21         |
| Phenology         | -0.149   | 0.085     | 1.758 | 0.079 | 15         |
| Max Altitude      | 0.147    | 0.084     | 1.746 | 0.081 | 25         |
| Range size        | 0.114    | 0.084     | 1.360 | 0.174 | 9          |
| PC2 ecophysiology | 0.042    | 0.084     | 0.505 | 0.613 | 8          |
| Wing size         | 0.029    | 0.068     | 0.428 | 0.669 | 6          |
| Host plants       | 0.026    | 0.072     | 0.357 | 0.721 | 4          |

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829 Table 3. Conditional average results among the selected models in a phylogenetic regression  
 830 comparing Dst diversity with butterfly traits. Prediction 2. Variables highlighted in bold showed a  
 831 significant effect.

| Dst               | Estimate      | Std.Error    | Z            | P            | models <sup>832</sup><br>(45) |
|-------------------|---------------|--------------|--------------|--------------|-------------------------------|
| <b>Phenology</b>  | <b>-0.206</b> | <b>0.081</b> | <b>2.547</b> | <b>0.011</b> | <del>833</del><br>23          |
| Range size        | 0.132         | 0.084        | 1.571        | 0.116        | <sup>834</sup><br>23          |
| PC1 ecophysiology | 0.122         | 0.084        | 1.460        | 0.144        | 19                            |
| Host plants       | -0.076        | 0.072        | 1.047        | 0.295        | 19                            |
| Max Altitude      | 0.094         | 0.084        | 1.125        | 0.261        | 15                            |
| Wing size         | -0.033        | 0.068        | 0.481        | 0.631        | 17                            |
| PC2 ecophysiology | 0.016         | 0.095        | 0.167        | 0.868        |                               |

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Table 4. Conditional average results among the selected models in a phylogenetic regression comparing Gst with butterfly traits. Prediction 2. Variables highlighted in bold showed a significant effect.

| Gst                | Estimate      | Std.Error    | Z            | P            | Models <sup>844</sup><br>(17) |
|--------------------|---------------|--------------|--------------|--------------|-------------------------------|
| <b>Host plants</b> | <b>-0.194</b> | <b>0.071</b> | <b>2.742</b> | <b>0.006</b> | <b>17</b>                     |
| <b>Range size</b>  | <b>0.195</b>  | <b>0.083</b> | <b>2.341</b> | <b>0.019</b> | <b>17</b>                     |
| <b>Wing size</b>   | <b>-0.135</b> | <b>0.067</b> | <b>2.025</b> | <b>0.043</b> | <b>12</b>                     |
| PC1 Phenology      | -0.141        | 0.074        | 1.922        | 0.055        | 11                            |
| Max Altitude       | -0.076        | 0.070        | 1.093        | 0.274        | 7                             |
| PC2 ecophysiology  | -0.061        | 0.093        | 0.655        | 0.512        | 6                             |
| PC1 ecophysiology  | 0.021         | 0.080        | 0.261        | 0.794        | 5                             |



Integrating three comprehensive datasets shows that mitochondrial DNA variation is linked to species traits and palaeogeographic events in European butterflies.

## Supplementary Methods and Results

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### ***DNA sequencing***

COI sequences generated for this study were obtained in two ways: In the first case, total genomic DNA was extracted using Chelex 100 resin, 100–200 mesh, sodium form (Biorad), under the following protocol: one leg was removed and introduced into 100 µl of Chelex 10% and 5 µl of Proteinase K (20 mg/ml) was added. The samples were incubated overnight at 55°C and were subsequently incubated at 100°C for 15 minutes. Samples were then centrifuged for 10 s at 3000 rpm. A 658-bp fragment near the 5' end of COI was amplified by polymerase chain reaction using the primers LepF1 and LepR1. Double-stranded DNA was amplified in 25-µL volume reactions containing: 14.4 µl autoclaved Milli-Q water, 5 µl 5x buffer, 2 µl 25 mM MgCl<sub>2</sub>, 0.5 µl 10 mM dNTPs, 0.5 µl of each primer (10 µM), 0.1 µl Taq DNA Polymerase (Promega, 5U/ µl) and 2 µl of extracted DNA. The typical thermal cycling profile followed this protocol: first denaturation at 92°C for 60 s, followed by five cycles of 92°C for 15 s, 48°C for 45 s and 62°C for 150 s, and then by 35 cycles of 92°C for 15 s, 52°C for 45 s and 62°C for 150 s and a final extension at 62°C for 420 s. PCR products were purified and sequenced by Macrogen Inc.

Other sequences were generated at the Biodiversity Institute of Ontario, Canada following standard protocols for DNA barcoding (deWaard, Ivanova, Hajibabaei, & Hebert, 2008), and DNA sequencing was performed on an ABI 3730XL capillary sequencer (Applied Biosystems).

Sequences were edited in CodonCode Aligner 3.0 or in GENEIOUS PRO 6.1.8 (Biomatters, <http://www.geneious.com/>) and assembled using the latter.

### ***D and G'st indexes***

Other than G<sub>st</sub> and D<sub>st</sub> we also computed D and G'st which are indicated as more reliable indicators of population differentiation when intra-area differentiation has particularly high values (which is usually not the case of mtDNA).

$$D = \frac{H_t - H_s}{1 - H_s} \times \frac{k}{k - 1}$$

$$G'st = \frac{\frac{H_t - H_s}{H_t} \times (k - 1 + H_s)}{(k - 1) \times (1 - H_s)}$$

where H<sub>t</sub> represents the average p-distances for all specimens of a given species, and H<sub>s</sub> is the average of the intra-unit p-distances and k represents the number of areas.

### ***Species recognized by Fauna Europaea lumped in the analysis***

COI differentiation shows a striking correspondence with butterfly taxonomy, but in some cases, species recognized as different taxa based on other markers (mostly morphological and ecological) share barcodes, thus underlying a shared mtDNA history. We considered these taxa as a single entity in our analyses. A review for the possibility to identify Western European species based on DNA barcoding is available in (Dapporto et al., 2017; Dincă et al., 2015; Vodă et al., 2016). Species lumped are listed below:

*Pyrgus alveus* (*P. alveus*, *P. accreta*, *P. bellieri*, *P. warrenensis*)

*Iphiclides podalirius* (*I. podalirius*, *I. feisthamelii*)

*Pieris napi* (*P. napi*, *P. bryoniae*)

*Phengaris alcon* (*P. alcon*, *P. rebeli*)

*Plebejus argus* (*P. argus*, *P. bellieri*)

*Plebejus idas* (*P. idas*, *P. argyrognomon*)

*Polyommatus dolus* (*P. dolus*, *P. ripartii*, *P. fulgens*, *P. fabressei*)

*Pseudophilotes baton* (*P. baton*, *P. vicrama*)

*Melitaea phoebe* (*M. phoebe*, *M. ornata*)

*Coenonympha arcania* (*C. arcania*, *C. darwiniana*, *C. gardetta*)

*Hipparchia semele* (*H. semele*, *H. blachieri*, *H. neapolitana*, *H. leighebi*, *H. sbordonii*)

*Erebia cassioides* (*E. cassioides*, *E. tyndarus*, *E. nivalis*, *E. calcaria*, *E. alvernensis*)

*Erebia ligea* (*E. ligea*, *E. euryale*)

*Erebia melampus* (*E. melampus*, *E. sudetica*)

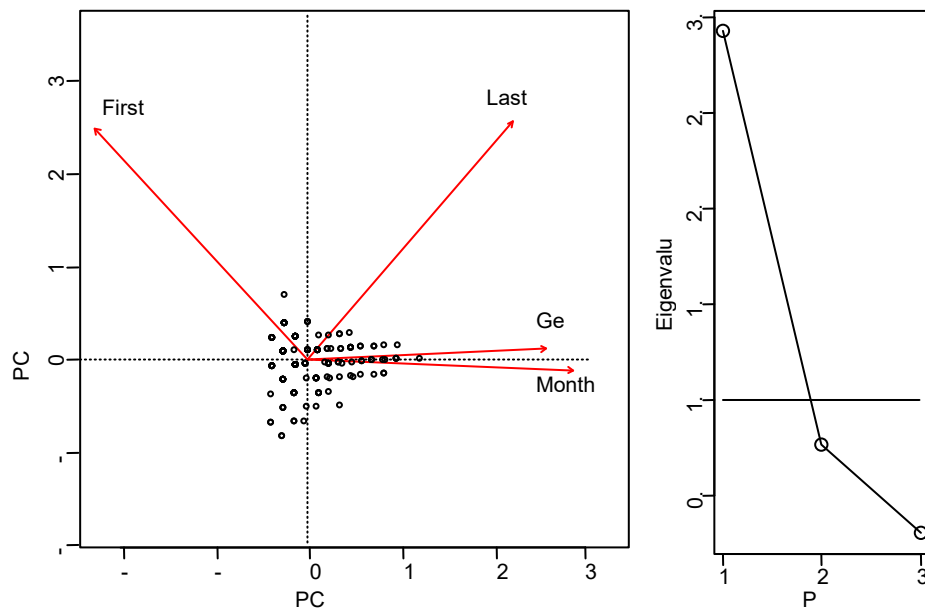


Fig S1. PCA scatterplot for phenology traits (left). First M, First month of emergence; Last M last month of emergence; Gen, number of generations; Months, length of flight period. The eigenvalue plot for the three components (right) shows that only the first PC had an eigenvalue higher than 1.

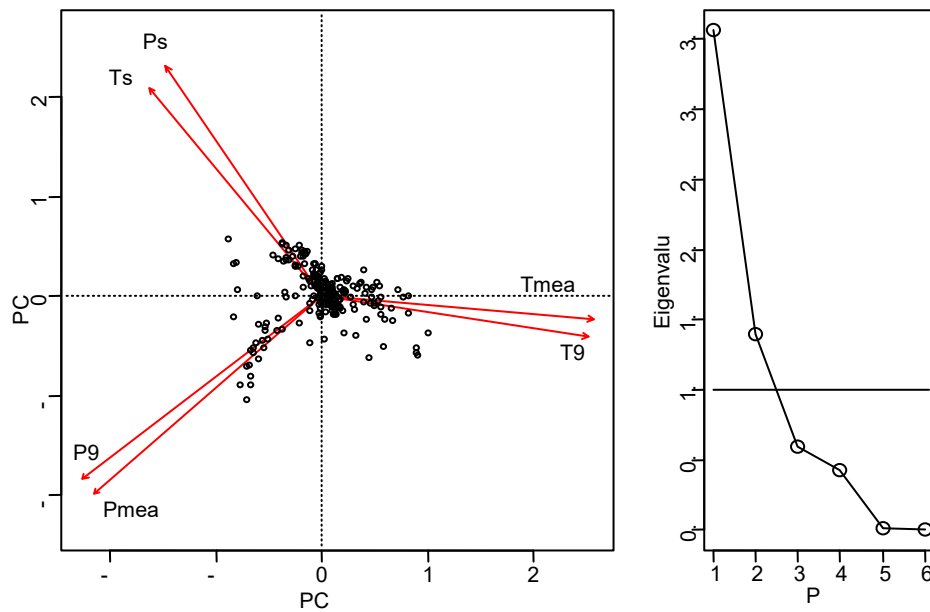


Fig S2. PCA scatterplot for ecophysiological traits (left). Tmean, mean temperature; Pmean, mean precipitation; T95, upper 95% confidence limit of temperature mean; P95, upper 95% confidence limit of precipitation mean; Tsd, standard deviation for temperature mean; Psd, standard deviation for precipitation mean. The eigenvalue plot for the six components (right) shows that the first two PCs had eigenvalues higher than 1.

Table S1.\_Conditional average results among the selected models in a phylogenetic regression comparing G'st with butterfly traits. Prediction 2.

| G'st               | Estimate      | Std.Error    | z            | P            | models(17) |
|--------------------|---------------|--------------|--------------|--------------|------------|
| <b>Range size</b>  | <b>0.195</b>  | <b>0.083</b> | <b>2.348</b> | <b>0.019</b> | <b>17</b>  |
| <b>Host plants</b> | <b>-0.194</b> | <b>0.071</b> | <b>2.734</b> | <b>0.006</b> | <b>17</b>  |
| <b>Wing size</b>   | <b>-0.135</b> | <b>0.067</b> | <b>2.023</b> | <b>0.043</b> | <b>12</b>  |
| Phenology          | 0.141         | 0.074        | 1.921        | 0.055        | 11         |
| Max Altitude       | -0.074        | 0.069        | 1.067        | 0.286        | 6          |
| PC2 ecophysiology  | -0.061        | 0.093        | 0.657        | 0.511        | 6          |
| PC1 ecophysiology  | 0.027         | 0.074        | 0.367        | 0.713        | 5          |

Table S2.\_Conditional average results among the selected models in a phylogenetic regression comparing D with butterfly traits. Prediction 2.

| D                 | Estimate     | Std.Error    | z            | P            | models(41) |
|-------------------|--------------|--------------|--------------|--------------|------------|
| <b>Phenology</b>  | <b>0.211</b> | <b>0.080</b> | <b>2.630</b> | <b>0.009</b> | <b>41</b>  |
| Max Altitude      | 0.099        | 0.084        | 1.174        | 0.240        | 17         |
| PC1 ecophysiology | 0.121        | 0.085        | 1.431        | 0.153        | 22         |
| Range size        | 0.111        | 0.082        | 1.351        | 0.177        | 20         |
| Host plants       | -0.079       | 0.072        | 1.104        | 0.270        | 18         |
| PC2 ecophysiology | 0.021        | 0.088        | 0.236        | 0.813        | 14         |
| Wing size         | -0.030       | 0.068        | 0.446        | 0.656        | 12         |

| Type of trait | Functional hypothesis  | Trait measured and description   | Sources  | PC1    | PC2    |
|---------------|--|--|--|--------|--------|
| Feeding       | Species feeding on a large number of plants have a higher potential to colonize new areas compared to species feeding on fewer plant species (Dennis et al., 2012)   | <b>Number of host plant genera</b> used by larvae as reported in two literature sources  | (Lafranchis, 2007)<br>(Tolman & Lewington, 2008) |        |        |
| Morphology    | Large-sized species are characterized by high mobility (Sekar, 2012) which increases the probability of crossing sea barriers (Dennis et al., 2012)  | <b>Wingspan</b> , calculated as the mean between minimum and maximum wing size reported in four main sources for European butterflies. | Higgins & Riley (1970)                           |        |        |
| Life history  | Phenological attributes characterize the period of the year and the duration of the most mobile life stage in butterflies, i.e. the winged adults. These characteristics can affect the probability of crossing sea barriers (Dapporto et al., 2012; Dennis et al., 2012) and can interact with climatic changes in determining extinction probabilities | <b>First month when adults fly</b> , ranging from January (1) to December (12)   | Tolman & Lewington (2008)                        | -0.456 | -      |
|               |  | <b>Last month when adults fly</b> , ranging from January (1) to December (12)  |  | 0.444  | -      |
|               |  | <b>Length of the flight period</b> : number of months when the adults occur in the study area  |  | 0.574  | -      |
|               |  | <b>Volitinism</b> : number of generations/year in the study area   |  | 0.515  | -      |
| Physiology    | Mean climatic conditions of the areas inhabited by a species are considered as good proxies for their ecophysiological response to climate (Devictor et al., 2012). They can affect the probability of species' persistence in the warm and dry Mediterranean climate that characterize the Tuscan islands   | <b>Mean temperature</b> occurring in the 50×50 km spatial cells where the species has been modelled to occur                           | (Schweiger, Harpke, Wiemers, & Settele, 2014)    | 0.448  | 0.347  |
|               |  | <b>Mean precipitation</b> in the same spatial cells as above   |  | -0.309 | 0.641  |
|               |  | <b>Maximum temperature tolerance</b> : upper 95% confidence interval for temperature mean  |  | 0.450  | 0.348  |
|               |  | <b>Minimum precipitation tolerance</b> : lower 95% confidence interval for precipitation mean  |  | -0.393 | 0.520  |
|               |  | <b>Overall temperature tolerance</b> : standard deviation for temperature mean   |  | -0.424 | -0.272 |
|               |  | <b>Overall precipitation tolerance</b> : standard deviation for precipitation mean   |  | -0.410 | 0.061  |
| Demography    | Distribution ranges and altitudinal distribution are expected to influence effective population size and the possibility for a species to diverge in different areas   | <b>Range size in Europe</b> : Number of 50×50 km spatial cells occupied in Europe  | (Schweiger, Harpke, Wiemers, & Settele, 2014)    | -      | -      |
|               |  | <b>Maximum Altitude</b> : Maximum altitude reported in Europe (m)  | Tolman & Lewington (2008)                        | -      | -      |
|               |  | <b>Altitudinal range</b> : Difference between minimum and maximum altitude reported in Europe  |  | -      | -      |

Table S3. Species traits used in the study with the description of the type of trait and the relative functional hypothesis; the trait(s) measured; the literature sources and the weights obtained by each trait in the first or the two first Principal Components (PC1 and PC2, when applied).

| Taxa                | COI        | ef1a       | wg         | RpS5       | GAPD<br>H  | CAD        | IDH       | MDH       | RPS2      | DDC       | HCL       | Thiola<br>se | CAT       | H3        |
|---------------------|------------|------------|------------|------------|------------|------------|-----------|-----------|-----------|-----------|-----------|--------------|-----------|-----------|
| <b>Hesperiidae</b>  | <b>47</b>  | <b>13</b>  | <b>15</b>  | <b>11</b>  | <b>8</b>   | <b>11</b>  | <b>11</b> | <b>11</b> | <b>5</b>  | <b>4</b>  | <b>3</b>  | <b>4</b>     | <b>4</b>  | <b>0</b>  |
| Hesperiinae         | 11         | 5          | 4          | 4          | 3          | 4          | 4         | 4         | 2         | 2         | 1         | 2            | 2         | 0         |
| Heteropterinae      | 3          | 3          | 3          | 3          | 2          | 3          | 3         | 3         | 1         | 1         | 1         | 1            | 1         | 0         |
| Pyrginae            | 33         | 5          | 8          | 4          | 3          | 4          | 4         | 4         | 2         | 1         | 1         | 1            | 1         | 0         |
| <b>Lycaenidae</b>   | <b>130</b> | <b>81</b>  | <b>64</b>  | <b>6</b>   | <b>9</b>   | <b>39</b>  | <b>6</b>  | <b>10</b> | <b>4</b>  | <b>3</b>  | <b>4</b>  | <b>4</b>     | <b>3</b>  | <b>55</b> |
| Aphnaeinae          | 1          | 1          | 1          | 1          | 1          | 1          | 1         | 1         | 1         | 1         | 1         | 1            | 1         | 1         |
| Lycaeninae          | 13         | 7          | 8          | 2          | 2          | 1          | 2         | 2         | 1         | 1         | 1         | 1            | 0         | 0         |
| Polyommatainae      | 99         | 66         | 53         | 1          | 5          | 35         | 1         | 5         | 1         | 0         | 1         | 1            | 1         | 53        |
| Theclinae           | 17         | 7          | 2          | 2          | 1          | 2          | 2         | 2         | 1         | 1         | 1         | 1            | 1         | 1         |
| <b>Nymphalidae</b>  | <b>246</b> | <b>147</b> | <b>173</b> | <b>109</b> | <b>105</b> | <b>34</b>  | <b>32</b> | <b>32</b> | <b>24</b> | <b>13</b> | <b>9</b>  | <b>9</b>     | <b>8</b>  | <b>1</b>  |
| Apaturinae          | 3          | 3          | 1          | 1          | 1          | 1          | 1         | 1         | 1         | 0         | 0         | 0            | 0         | 0         |
| Charaxinae          | 1          | 1          | 1          | 1          | 0          | 0          | 0         | 0         | 1         | 0         | 0         | 0            | 0         | 0         |
| Danainae            | 2          | 2          | 2          | 1          | 1          | 1          | 1         | 2         | 2         | 1         | 0         | 0            | 0         | 0         |
| Heliconiinae        | 32         | 31         | 31         | 12         | 13         | 4          | 3         | 2         | 5         | 0         | 0         | 0            | 0         | 0         |
| Libytheinae         | 1          | 1          | 1          | 1          | 1          | 1          | 1         | 1         | 1         | 1         | 0         | 0            | 0         | 0         |
| Limenitidinae       | 5          | 5          | 5          | 4          | 5          | 2          | 5         | 3         | 2         | 2         | 1         | 1            | 1         | 0         |
| Nymphalinae         | 37         | 33         | 32         | 19         | 16         | 14         | 10        | 12        | 4         | 3         | 2         | 2            | 2         | 1         |
| Satyrinae           | 165        | 71         | 100        | 70         | 68         | 11         | 11        | 11        | 8         | 6         | 6         | 6            | 5         | 0         |
| <b>Papilionidae</b> | <b>15</b>  | <b>14</b>  | <b>12</b>  | <b>3</b>   | <b>2</b>   | <b>3</b>   | <b>3</b>  | <b>2</b>  | <b>2</b>  | <b>2</b>  | <b>2</b>  | <b>1</b>     | <b>2</b>  | <b>0</b>  |
| Papilioninae        | 5          | 5          | 3          | 0          | 0          | 0          | 0         | 0         | 0         | 1         | 0         | 0            | 0         | 0         |
| Parnassiinae        | 10         | 9          | 9          | 3          | 2          | 3          | 3         | 2         | 2         | 1         | 2         | 1            | 2         | 0         |
| <b>Pieridae</b>     | <b>57</b>  | <b>26</b>  | <b>18</b>  | <b>13</b>  | <b>13</b>  | <b>15</b>  | <b>12</b> | <b>11</b> | <b>6</b>  | <b>4</b>  | <b>3</b>  | <b>3</b>     | <b>3</b>  | <b>0</b>  |
| Coliadinae          | 18         | 8          | 4          | 3          | 3          | 3          | 3         | 2         | 3         | 1         | 1         | 1            | 1         | 0         |
| Dismorphiinae       | 5          | 3          | 5          | 2          | 2          | 4          | 2         | 2         | 1         | 1         | 1         | 1            | 1         | 0         |
| Pierinae            | 34         | 15         | 9          | 8          | 8          | 8          | 7         | 7         | 2         | 2         | 1         | 1            | 1         | 0         |
| <b>Riodinidae</b>   | <b>1</b>   | <b>1</b>   | <b>1</b>   | <b>1</b>   | <b>0</b>   | <b>1</b>   | <b>1</b>  | <b>1</b>  | <b>1</b>  | <b>1</b>  | <b>0</b>  | <b>0</b>     | <b>0</b>  | <b>1</b>  |
| Nemeobiinae         | 1          | 1          | 1          | 1          | 0          | 1          | 1         | 1         | 1         | 1         | 0         | 0            | 0         | 1         |
| <b>TOTAL</b>        | <b>496</b> | <b>282</b> | <b>283</b> | <b>143</b> | <b>137</b> | <b>103</b> | <b>65</b> | <b>67</b> | <b>42</b> | <b>27</b> | <b>21</b> | <b>21</b>    | <b>20</b> | <b>57</b> |
| Coverage            | 100%       | 57%        | 57%        | 29%        | 28%        | 21%        | 13%       | 14%       | 8%        | 5%        | 4%        | 4%           | 4%        | 11%       |

Table S4. Coverage for different mitochondrial and nuclear markers for the sequence data used to construct the phylogenetic tree of the European butterflies.

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